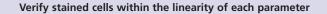


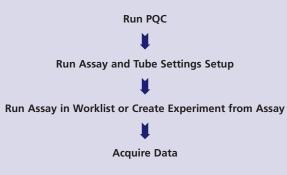
Determine 2.5 x rSD<sub>ru</sub> with unstained cells



Save optimal settings and create an assay

Transfer the assay to remaining cytometers

# **Reusing optimal settings in BD FACSuite software**



Note: If reference settings were created during the initial optimization, they will need to be updated every 30 days.

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**BD FACSuite<sup>™</sup> Software** 

Standardizing Application Settings Across Multiple Cytometers

Multicolor flow cytometry is increasingly used in research laboratories to generate more data from a given experiment.

A key aspect of effectively using multicolor flow is ensuring that the experiment can be performed consistently over time and across instruments. Standardizing experiments using application setup can ensure consistency of results over time across multiple systems. To achieve this consistency, we recommend creating and transferring Assays in BD FACSuite software.

To create and save the optimal settings for specific experiments, follow these basic steps:

> Step 1: Run Characterization and Performance QC Step 2: Determine 2.5 x rSD<sub>EN</sub> with unstained cells Step 3: Verify stained cells within the linearity of each parameter

Step 4: Save optimal settings and create an assay Step 5: Transfer the assay to remaining cytometers



### Step 1: Run Characterization QC (CQC) and Performance QC (PQC)

- a. Verify that CQC has been run.
- b. Run PQC on each cytometer.

Name

Reference Particles:P1

c. Record the rSD<sub>EN</sub> and linearity max for each fluorescence channel for each cytometer.

### Step 2: Determine 2.5 x rSD<sub>EN</sub> with unstained cells

- a. Multiply the  $rSD_{EN}$  by 2.5 for all fluorescence channels.
- b. Compare these values across all cytometers and record the highest value for each channel.

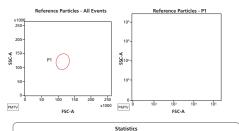
FITC-A PE-A PerCP-Cy5.5-A PE-Cy7-A APC-A APC-Cy7-A V450-A

\*\*\* \*\*\*

RSD

RSD RSD RSD RSD RSD RSD

c. Adjust the PMT voltage (PMTV) for each channel until the rSD for unstained cells reaches this value.

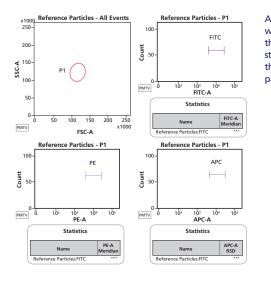


An example of a worksheet used to acquire unstained cells and adjust PMTVs until the target rSD values (2.5 x rSD<sub>EN</sub>) are reached.

# Step 3: Verify stained cells within the linearity of each parameter

- a. Divide the linearity max value by 2 for all fluorescence channels.
- b. Compare these values across all cytometers and record the lowest value for each channel.
- c. Using highly positive cell populations, verify that the median fluorescence intensity (MFI) of the positive population falls at or below this number.

Note: If the MFI exceeds this value for any channel, lower the PMTV.



An example of a worksheet used to verify that MFI values for stained cells fall within the linearity of each parameter.

### Step 4: Save optimal settings and create an Assay

a. Create tube settings or reference settings by right-clicking the tube and selecting Create Tube Settings or Create Reference Settings. b. Create an assay by selecting **File** > **Create Assay**.

#### Step 5: Transfer settings to the remaining cytometers

- a. Export your assay from the first cytometer. In the **User-Defined Assays** section of the Library, select your assay, then click File > Export.
- b. Import your assay to the remaining cytometers. In the User-Defined **Assays** section of the Library, select **File** > **Import**.
- c. Run assay and tube setting setup for the appropriate assay. In the Setup & QC workspace, select your assay from the Assay & Tube Settings Setup task menu, then click Start.